

## ○ Polymeric partition chromatography columns and materials MCI GEL™ CHP series

### Separation mechanism of CHP series

High performance liquid chromatography relies on one of the following physical phenomena for efficient separation of solutes: partition, adsorption, size exclusion, or ion exchange. Of these, partition chromatography is the most commonly used method, and it separates solutes based on their difference in partitioning between a stationary phase and a mobile phase. This technique has currently become the mainstay in industry for the separation of organic compounds such as pharmaceuticals, agricultural chemicals, and other intermediates. Practically, partition chromatography can be performed in two different modes depending on the relative polarities of the stationary and mobile phases. In the normal phase (NP) mode, the mobile phase is less polar than the stationary phase while the situation is reversed in the reverse phase (RP) mode, where the mobile phase is significantly more polar than the stationary phase.

MCI GEL™ specializes in polymer-based packing materials. The use of polymer-based columns has become more widespread thanks to the many advantages of the polymer matrix like excellent selectivity, the absence of specific adsorption which is found commonly with silica-based packing, operability in a wide pH range and good chemical stability due to the inert nature of polymeric materials. The MCI GEL™ partition chromatography columns are based on a polystyrene and polymethacrylate porous polymer. As RP columns, they are applied to the separation of a wide variety of organic compounds, both in the isocratic and gradient elution mode. The compounds include peptides, insulin, small molecule APIs, nutraceutical compounds, water-soluble vitamins and nucleotides. As NP columns, they are used in the separation of various carotenoids, fat-soluble vitamins, steroids, and food additives. These columns tolerate various organic solvents like hexane, heptane, methylene chloride, and alcohols.

As NP columns, they are used in the separation of various carotenoids, fat-soluble vitamins, steroids, and food additives. Various organic solvents like Hexane Heptane, methylene chloride and alcohols can be used.

The MCI GEL™ packing materials are based on the same chemistries offered in the Diaion™ and Sepabeads™ synthetic adsorbent resins. These polymer chemistries, like Diaion™ HP series and Sepabeads™ SP series, are widely used and documented in the biopharmaceutical industry for fermentation extraction, the food industry and in industrial chromatographic separations. The MCI GEL™ packing materials are available as packed columns for analytical applications, and as bulk packing materials for analytical, preparative and production chromatography applications.

#### ● Description of MCI GEL™ columns and materials

### MCI GEL™ CHP20/C04

Matrix type

Particle size

{ C=Column  
P=Material

MCI GEL™ CHP series are suitable for RP and NP chromatography. There are four kinds of columns of various hydrophobicities; porous polystyrene, modified porous polystyrene, polymethacrylate, and modified porous polymethacrylate. This range of packing materials offers tremendous scope for a proper selection of columns based on the properties of the target compounds.

Polystyrene packing: MCI GEL™ CHP20/C04, CHP20/C10

Modified polystyrene packing: MCI GEL™ CHP07/C04, CHP07/C10, CHK40/C04

Polymethacrylate packing: MCI GEL™ CMG20/C10

Modified polymethacrylate packing: MCI GEL™ CHPOD/C04, CHK45/C05

The hydrophobicities of the columns are in the following orders:

MCI GEL™ CHP07/C04, C10 > CHP20/C04, C10 > CHPOD/C04 ≥ ODS columns ≥ CMG20/C04, C10

Polymer columns for HPLC, with their superior chemical resistance, can be used with various mobile phases of broad pH range, acidic through alkaline. They have the following advantages due to their high hydrophobicities:

- 1) In reverse phase chromatographic methods to separate acidic or alkaline compounds, eluents that can suppress the ionic properties of such compounds are generally used. Polymer columns can be applied in these cases where ODS columns would be unsuitable.
- 2) Some extremely hydrophilic compounds, e.g., oligosaccharides, can be separated using strongly hydrophobic CHP07/C04 or CHP07/C10 columns.
- 3) Polymer columns can be washed with acidic and/or basic solutions in case of contamination.

Polymethacrylate columns, CMG20/C04 and CMG20/C10, can be applied both for reverse phase and normal phase chromatography.

Modified polystyrene packing, CHK40/C04, is a mixed-mode type material; both hydrophobic and hydrophilic interactions occur between the packing material surface and the analytes. This material is useful for compounds that are difficult to separate using existing ODS or other polymer-based columns. This column is also used in the normal phase mode and shows a unique separation profile.

All polymeric columns exhibit superior stability and yield in comparison to ODS columns, which may have free silanol groups even when end-capping agents have been used.

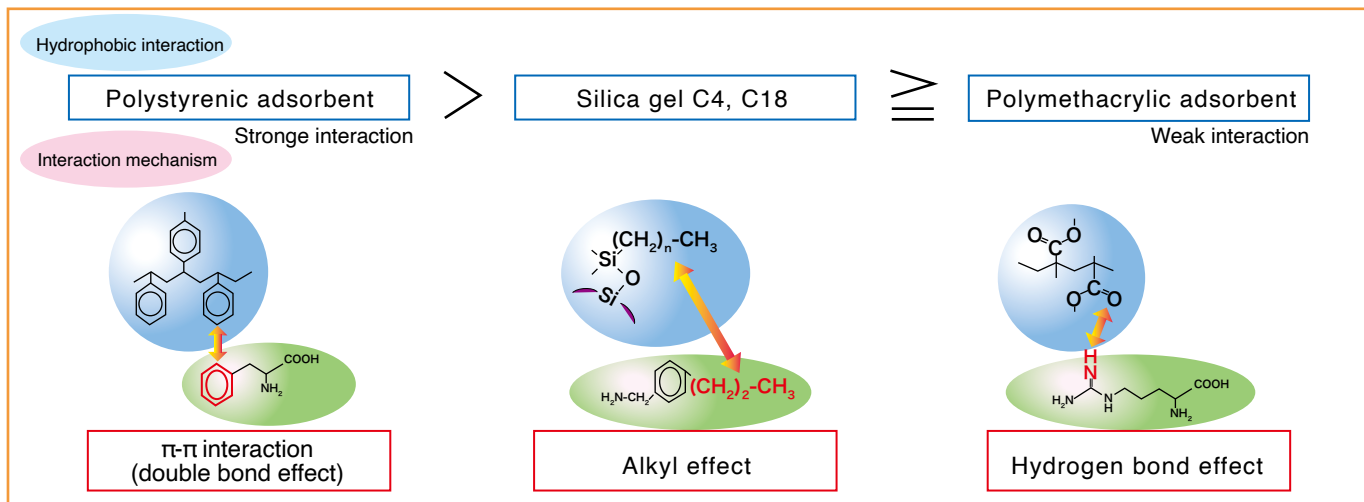
## Column list

### ● CHP column series

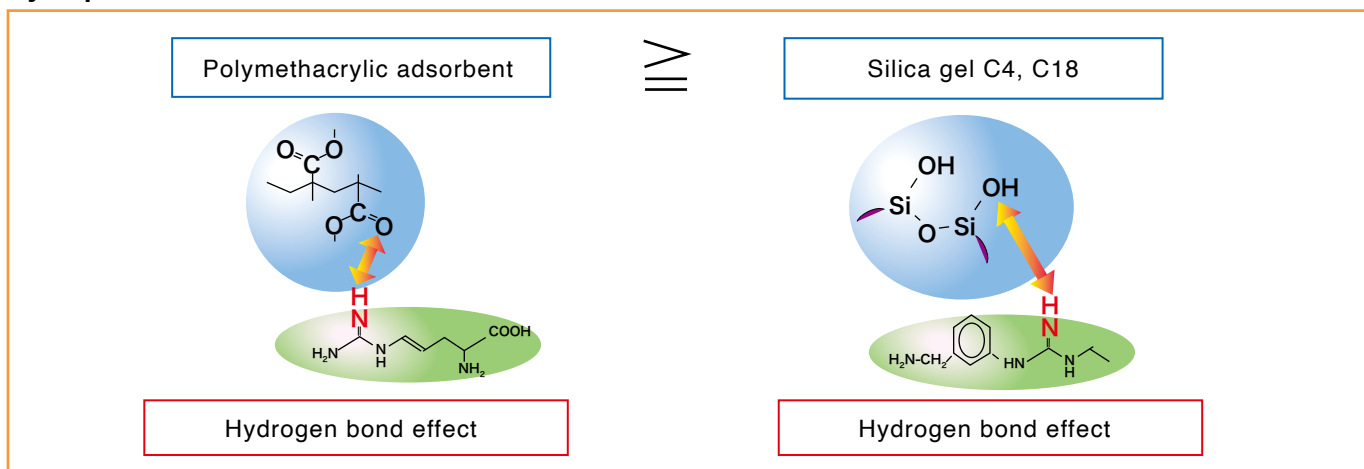
Matrix Type	Functional group	Product name	Particle size [μm]	Column size [mm I.D.×mm]	pH range	USP
Styrene Divinylbenzene	None	CHP20/C04	4	4.6×150 20×150	Full range	L21
		CHP20/C10	10	4.6×150 4.6×250 10×250 20×150 20×250		
	Br	CHP07/C04	4	4.6×150 20×200		
		CHP07/C10	10	4.6×150 4.6×250 10×150 20×150 20×250		
	Cation exchange group	CHK40/C04	4	4.6×150		
Methacrylates	None	CMG20/C04	4	4.6×150 20×150	2~12	
		CMG20/C10	10	4.6×150 4.6×250 10×250 20×150 20×250		
	C18	CHPOD/C04	4	4.6×150 20×200		
	Weak cation exchange group	CHK45/C05	5	4.6×150		

\*CHP20/C04, CHP20/C10: USP classification is L21

### Retentiveness in reverse phase mode



### Hydrophobic interaction Interaction mechanism

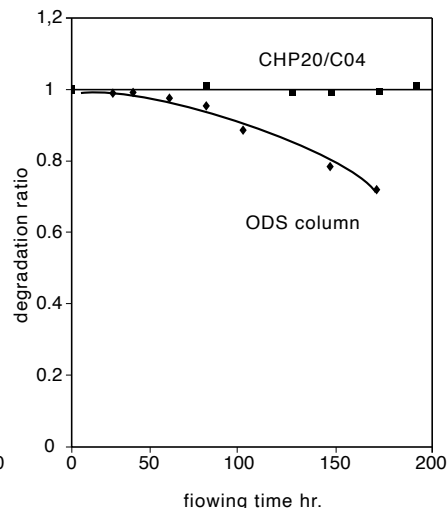
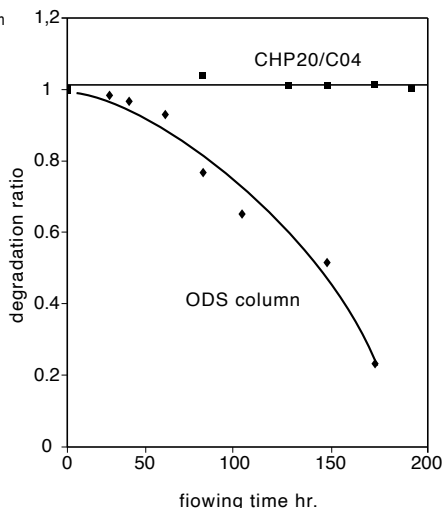


## Durability of polymeric column

The polymeric RP columns are chemically stable. Specifically, the columns have resistance to an alkaline eluent. The following graphs demonstrate stability of the polymeric columns. After feeding a solution of pH 12 into the MCI GEL™ CHP20/C04, there is no change of column performance.

Fig. 5-1 Column durability at pH12 comparison between CHP20/C04 and an ODS column

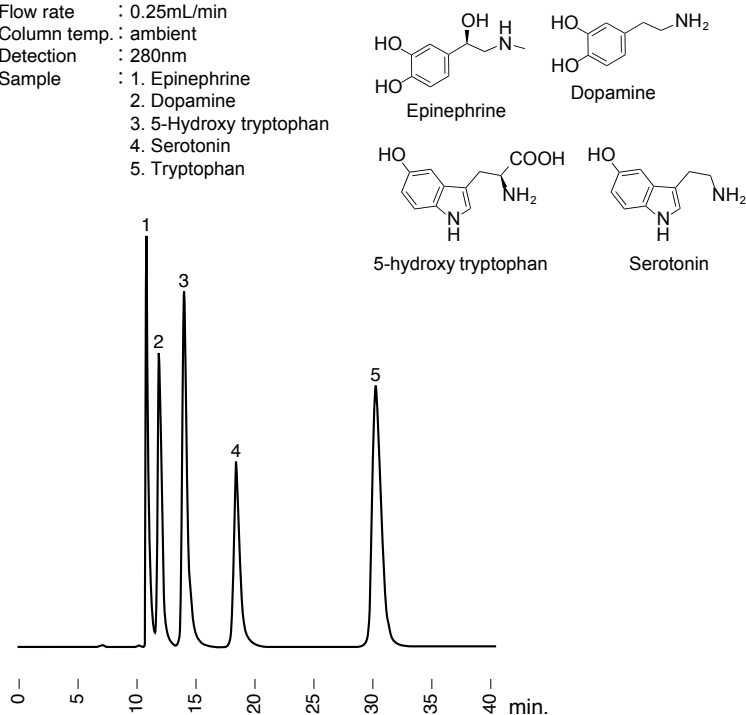
Conditions  
 Column : MCI GEL™ CHP20/C04 4.6mm.I.D × 150mm  
 Eluent : 20mM Na<sub>2</sub>HPO<sub>4</sub> pH12/CH<sub>3</sub>CN/=60/40  
 Flow rate : 0.4mL/min  
 Column temp.: 25°C  
 Detection : 254nm  
 Sample : 1000ppm Dimethyl phthalate 5μL



# Application data of CHP series

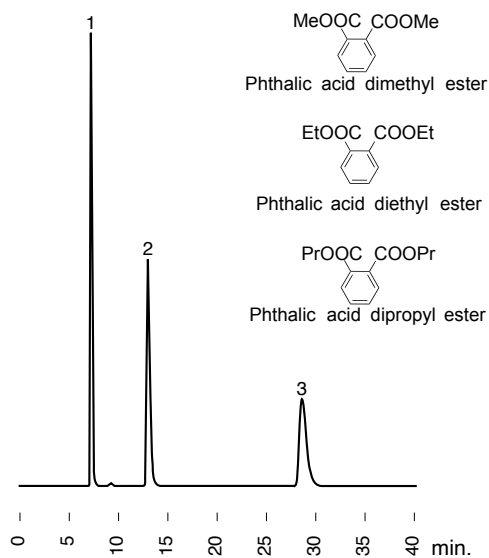
**Fig. 5-2 Separation of catecholamines**

Conditions  
 Column : MCI GEL™ CHP20/C04  
 4.6mm I.D.×150mm  
 Eluent : 50mM Na-phosphate pH2.0,  
 1.5% Hexanesulfonic acid /  
 CH<sub>3</sub>CN=80/20  
 Flow rate : 0.25mL/min  
 Column temp. : ambient  
 Detection : 280nm  
 Sample : 1. Epinephrine  
 2. Dopamine  
 3. 5-Hydroxy tryptophan  
 4. Serotonin  
 5. Tryptophan



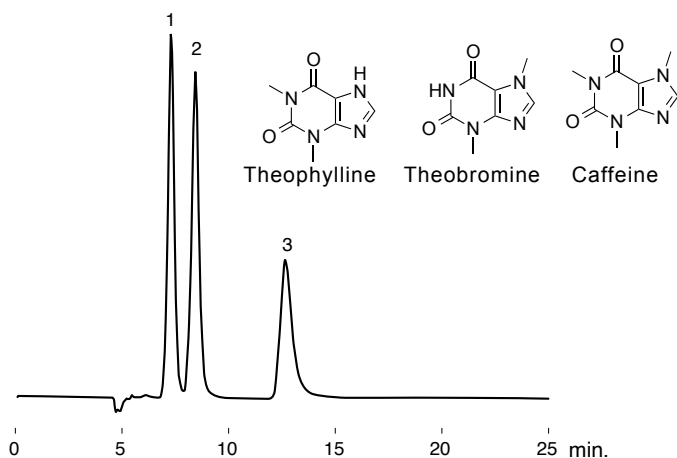
**Fig. 5-3 Separation of phthalic acid esters**

Conditions  
 Column : MCI GEL™ CHP20/C04  
 4.6mm I.D.×150mm  
 Eluent : H<sub>2</sub>O/CH<sub>3</sub>CN=50/50  
 Flow rate : 0.75mL/min  
 Column temp. : 60°C  
 Detection : 254nm  
 Sample : 1. Dimethyl phthalate  
 2. Diethyl phthalate  
 3. Dipropyl phthalate



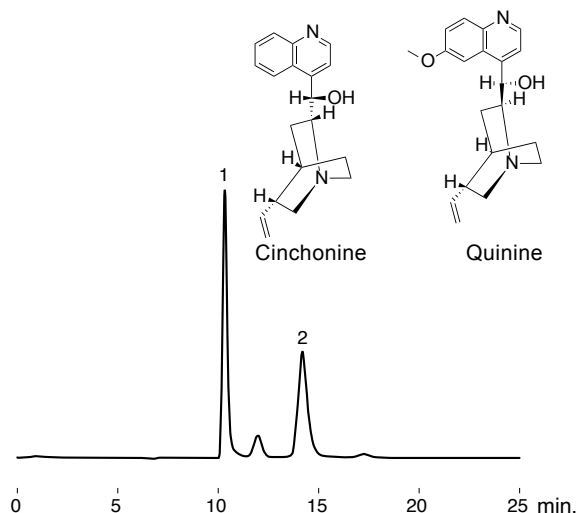
**Fig. 5-4 Purine alkaloids**

Conditions  
 Column : MCI GEL™ CHP20/C04  
 4.6mm I.D.×150mm  
 Eluent : H<sub>2</sub>O/CH<sub>3</sub>CN=10/90  
 Flow rate : 0.4mL/min  
 Column temp. : 25°C  
 Detection : 275nm  
 Sample : 1.Theophylline  
 2.Theobromine  
 3.Caffeine



**Fig. 5-5 Cinchona alkaloids**

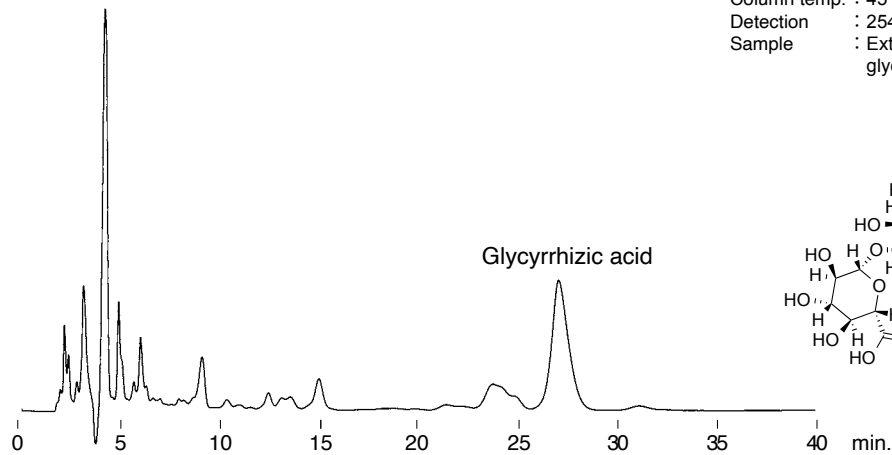
Conditions  
 Column : MCI GEL™ CHP20/C04  
 4.6mm I.D.×150mm  
 Eluent : 0.1M NaH<sub>2</sub>PO<sub>4</sub> pH2.0  
 CH<sub>3</sub>CN=88/12  
 Flow rate : 0.3mL/min  
 Column temp. : 25°C  
 Detection : 275nm  
 Sample : 1.Cinchonine  
 2.Quinine



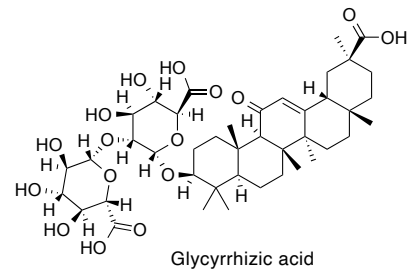


# Application data of CHP series

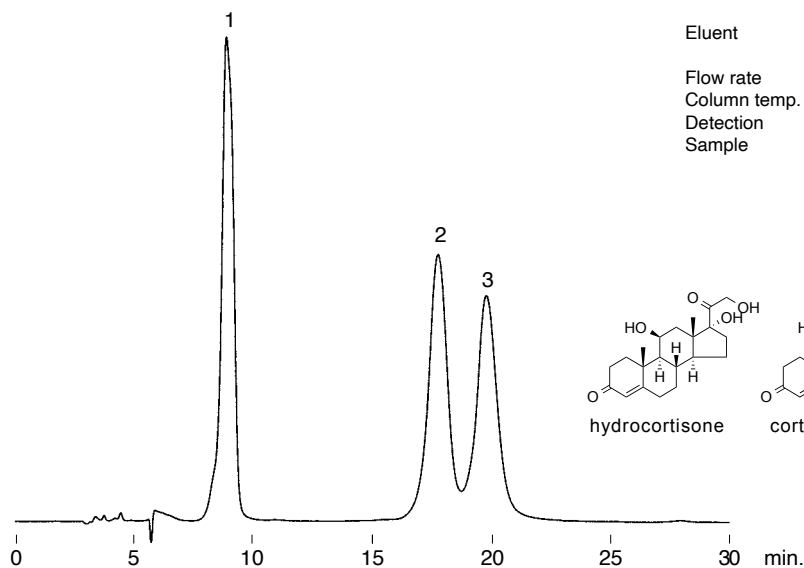
**Fig. 5-8 Glycyrrhizae radix**



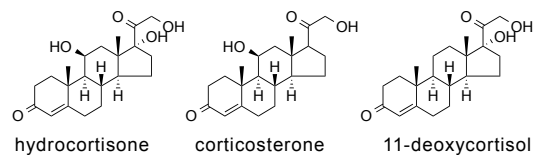
Conditions  
 Column : MCI GEL™ CHP20/C04  
 4.6mm I.D.×150mm  
 Eluent : 2.06% acetic acid/CH<sub>3</sub>CN  
 =63/37  
 Flow rate : 0.5mL/min  
 Column temp. : 45°C  
 Detection : 254nm  
 Sample : Extract of  
 glycyrrhizae radix



**Fig. 5-9 Adrenal cortex hormones**

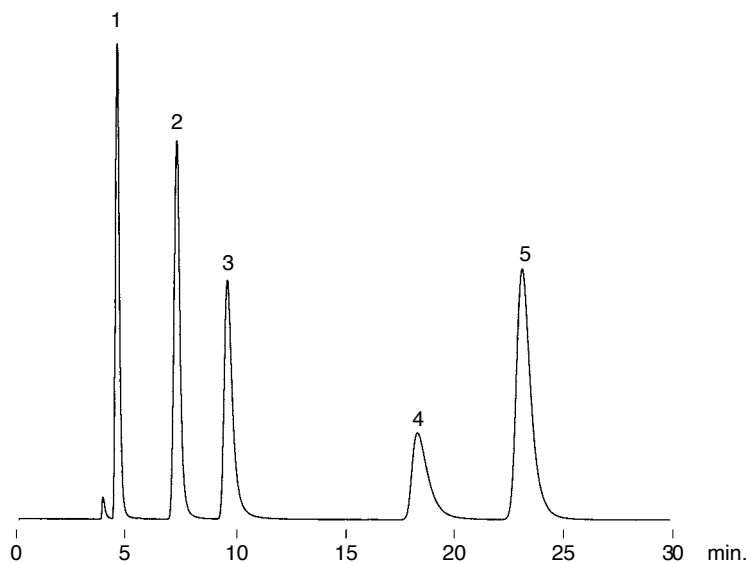


Conditions  
 Column : MCI GEL™ CHP20/C04  
 4.6mm I.D.×150mm  
 Eluent : H<sub>2</sub>O/CH<sub>3</sub>CN  
 : 60/40  
 Flow rate : 0.5mL/min  
 Column temp. : 45°C  
 Detection : 280nm  
 Sample : 1. Hydrocortisone  
 2. Corticosterone  
 3. 11-Deoxycortisol



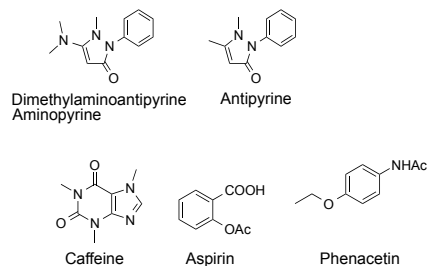
## Application data of CHP series

Fig. 5-10 Ingredients of medicine



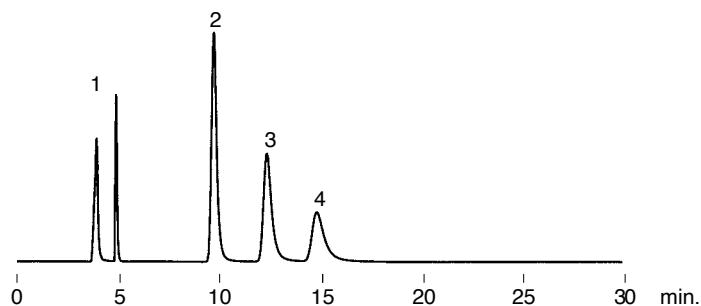
Conditions

Column : MCI GEL™ CMG20/C04  
4.6mm I.D.×150mm  
Eluent : 50mM phosphoric acid(pH2.0)/CH<sub>3</sub>OH  
=60/40  
Flow rate : 0.5mL/min  
Column temp. : 45°C  
Detection : 254nm  
Sample : 1.4-Dimethylaminoantipyrine  
2.Antipyrine  
3.Caffeine  
4.Aspirin  
5.Phenacetin



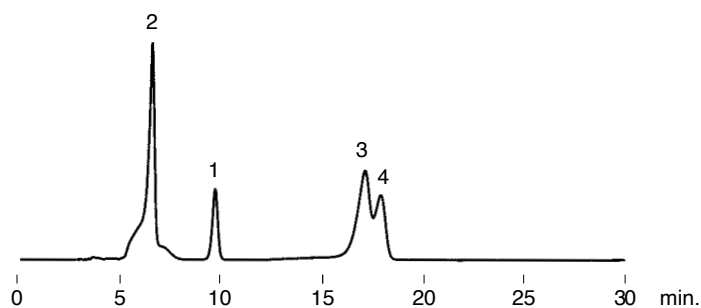
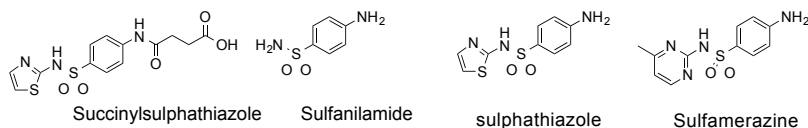
## Comparison with an ODS column

Fig. 5-11 Sulfa drugs



Conditions

Column : MCI GEL™ CMG20/C04  
4.6mm I.D.×150mm  
Eluent : 20mM phosphate pH6.8/CH<sub>3</sub>CN  
=82/18  
Flow rate : 0.5mL/min  
Column temp. : 45°C  
Detection : 254nm  
Sample : 1.Succinylsulfathiazole  
2.Sulfanilamide  
3.Sulfathiazole  
4.Sulfamerazine



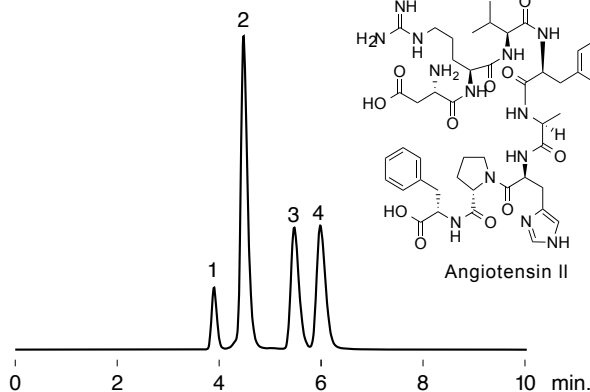
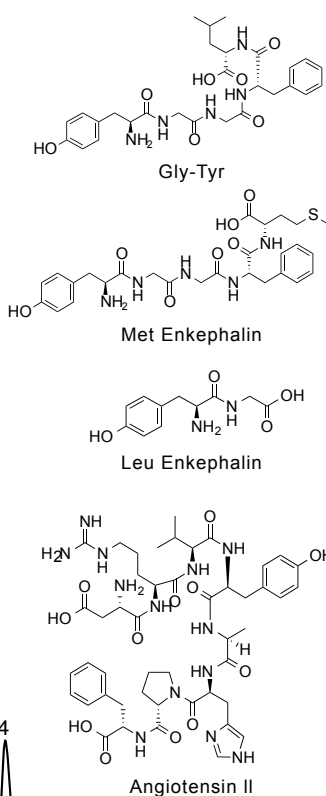
Conditions

Column : ODS column  
4.6mm I.D.×150mm  
Eluent : 20mM phosphate pH6.8/CH<sub>3</sub>CN  
=90/10  
Flow rate : 0.5mL/min  
Column temp. : 45°C  
Detection : 254nm  
Sample : 1.Succinylsulfathiazole  
2.Sulfanilamide  
3.Sulfathiazole  
4.Sulfamerazine

# Application data of CHP series

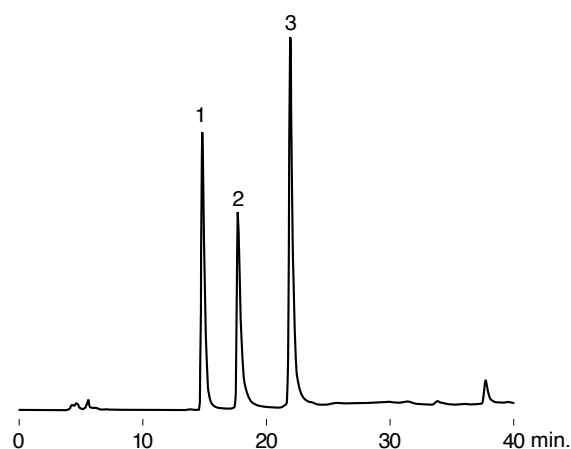
**Fig. 5-12 Peptides**

Conditions  
 Column : MCI GEL™ CMG20/C04  
 4.6mm I.D.×150mm  
 Eluent : 0.1%TFA/CH<sub>3</sub>CN  
 =70/30  
 Flow rate : 0.5mL/min  
 Column temp. : 25°C  
 Detection : 220nm  
 Sample : 1.Gly-Tyr  
 2.Met Enkephalin  
 3.Leu Enkephalin  
 4.Angiotensin II



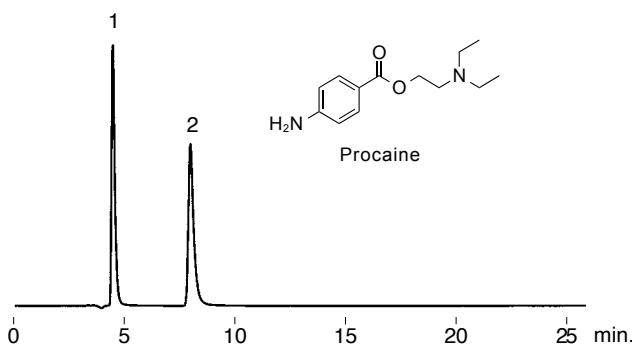
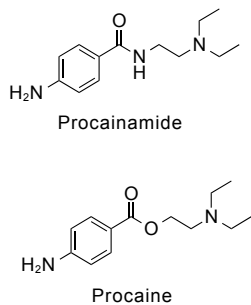
**Fig. 5-13 Proteins**

Conditions  
 Column : MCI GEL™ CMG20/C04  
 4.6mm I.D.×150mm  
 Eluent : A;0.05%TFA/CH<sub>3</sub>CN  
 =80/20  
 B;0.05%TFA/CH<sub>3</sub>CN  
 =20/80  
 A→B 30min.linear  
 Flow rate : 0.5mL/min  
 Column temp. : 25°C  
 Detection : 280nm  
 Sample : 1.Ribonuclease A  
 2.Cytochrome c  
 3.α-Chymotrypsinogen A



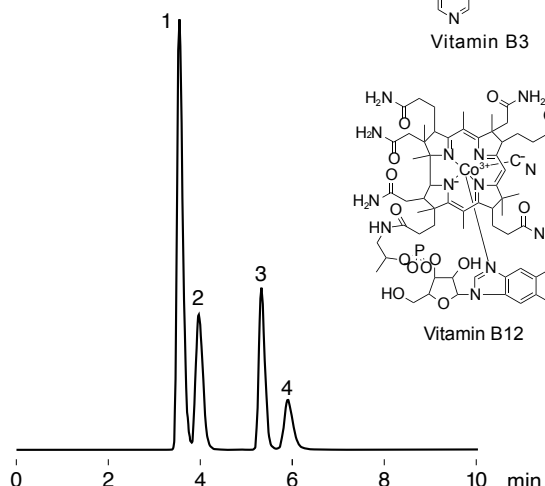
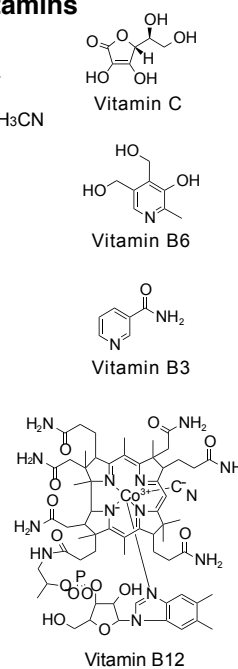
**Fig. 5-14 Procainamide, Procaine**

Conditions  
 Column : MCI GEL™ CMG20/C04  
 4.6mm I.D.×150mm  
 Eluent : 20mM phosphate pH7.2/CH<sub>3</sub>CN  
 =65/35  
 Flow rate : 0.5mL/min  
 Column temp. : 45°C  
 Detection : 254nm  
 Sample : 1.Procainamide  
 2.Procaine



**Fig. 5-15 Water-soluble vitamins**

Conditions  
 Column : MCI GEL™ CMG20/C04  
 4.6mm I.D.×150mm  
 Eluent : 8mM Na<sub>2</sub>HPO<sub>4</sub> pH7.0/CH<sub>3</sub>CN  
 =85/15  
 Flow rate : 0.5mL/min  
 Column temp. : 25°C  
 Detection : 254nm  
 Sample : 1.Vitamin C  
 2.Vitamin B6  
 3.Vitamin B3  
 4.Vitamin B12

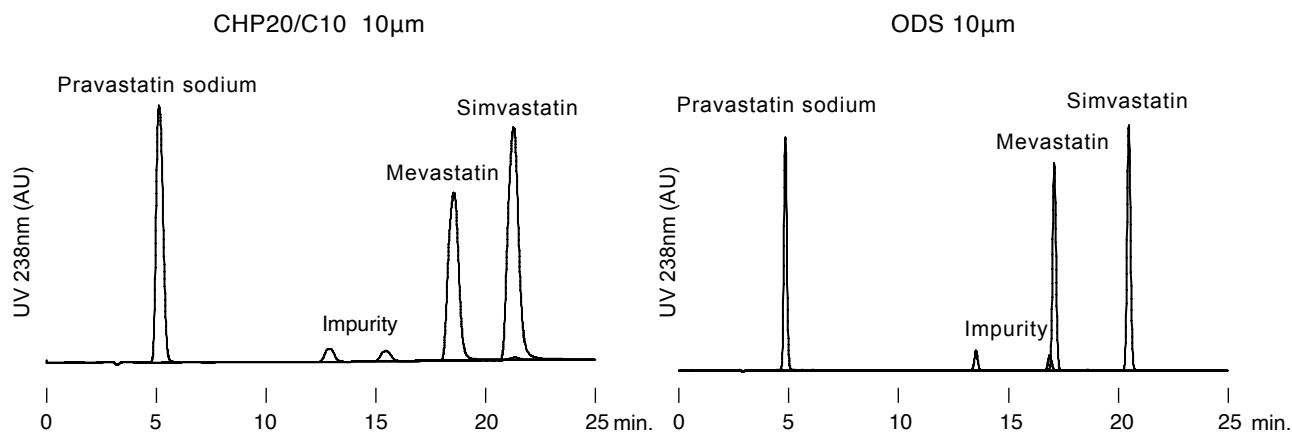
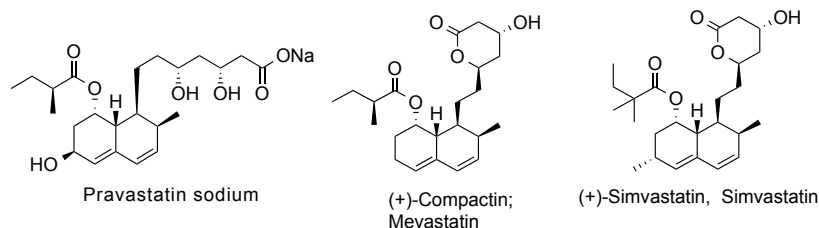




# Application data of CHP series

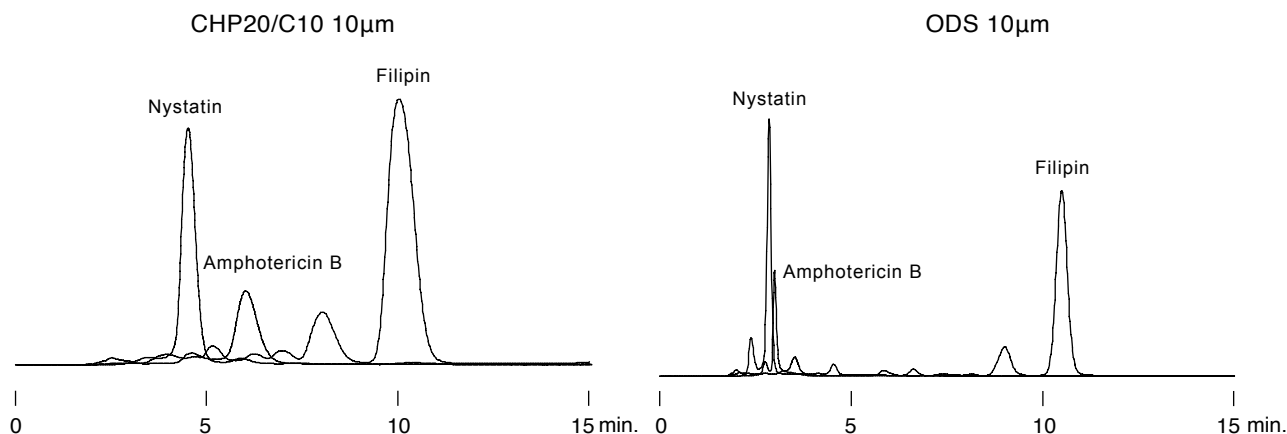
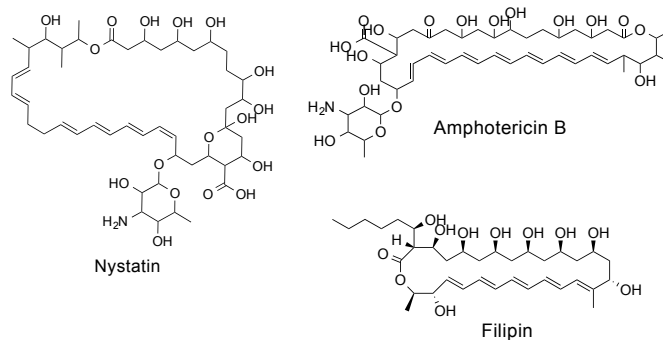
**Fig. 5-16 Pravastatin sodium**

**Conditions**  
**Column** : MCI GEL™ CHP20/C10 (10µm 250 ×4.6mm I.D.) and ODS (10µm 250 ×4.6mm I.D.)  
**Eluent** : A :0.1% Formic acid; B :0.1% Formic acid in AcCN;  
**Gradient** : 45%B-95%B over 29min.  
**Flow rate** : 1.00mL/min  
**Column temp.** : 25°C  
**Detection** : UV238nm  
**Sample** : Pravastatin sodium, Mevastatin and Simvastatin, 1mg/ml each;.  
**Injection** : 5µL



**Fig. 5-17 Polyene antibiotics**

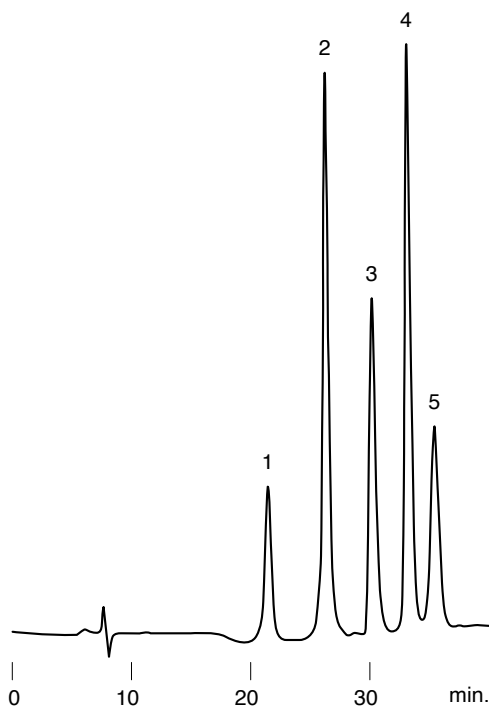
**Conditions**  
**Column** : MCI GEL™ CHP20/C10 (10µm 250 ×4.6mm I.D.) and ODS (10µm 250 ×4.6mm I.D.)  
**Eluent** : A :0.1% Formic acid; B :0.1% Formic acid in AcCN; A/B=60/40;  
**Flow rate** : 1.00mL/min  
**Column temp.** : 25°C  
**Detection** : UV305nm for Nystatin, VIS405nm for Amphotericin B and UV340nm for Filipin;  
**Sample** : Pravastatin sodium, Mevastatin and Simvastatin, 1mg/ml each;.  
**Injection** : 10µL



# Application data of CHP series

**Fig. 5-18 Proteins**

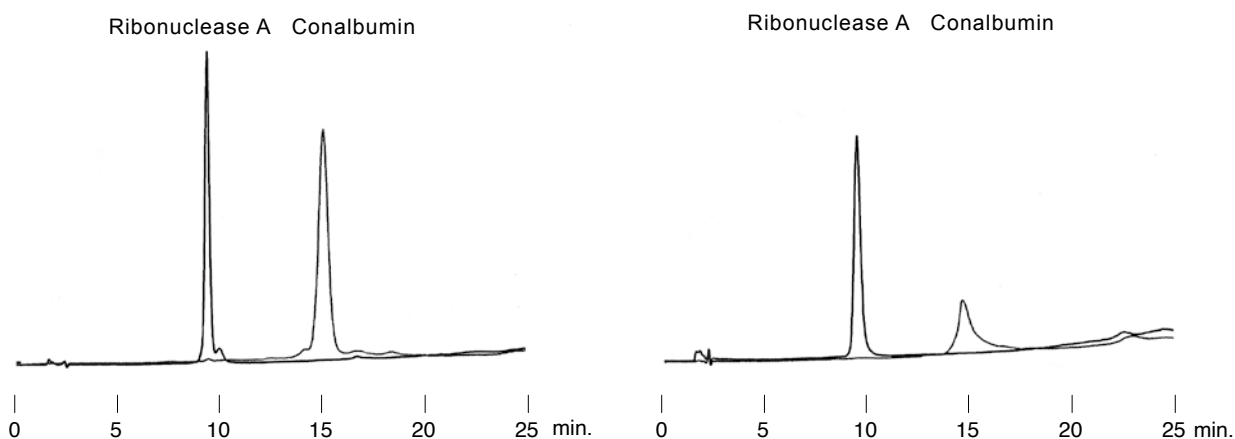
Conditions  
 Column : MCI GEL™ CMG20/C10  
 4.6mm I.D.×250mm  
 Eluent : A 0.05% TFA/CH<sub>3</sub>CN=80/20  
 B 0.05% TFA/CH<sub>3</sub>CN=30/70  
 A → B 45min linear gradient  
 Flow rate : 0.5mL/min  
 Column temp. : 25°C  
 Detection : 280nm  
 Sample : 1. Ribonuclease A  
 2. Cytochrome C  
 3. Transferrin  
 4. α-Chymotrypsinogen A  
 5. β-Lactoglobulin



**Fig. 5-19 Proteins**

CHP20/C10 10μm

ODS 10μm

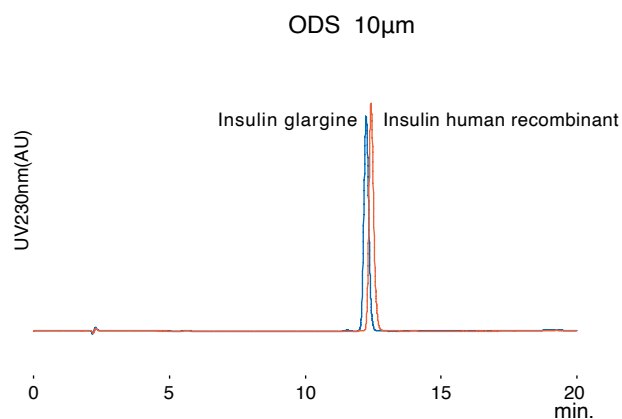
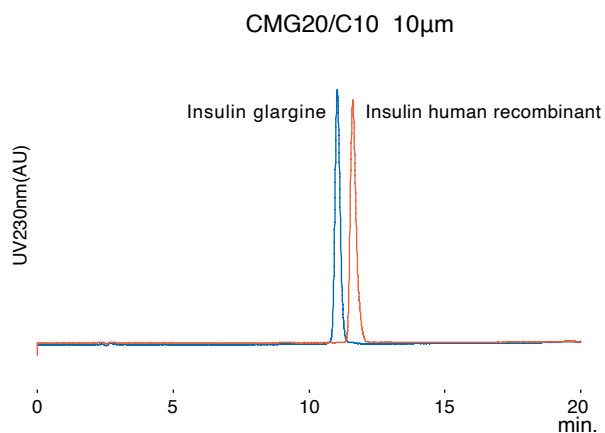
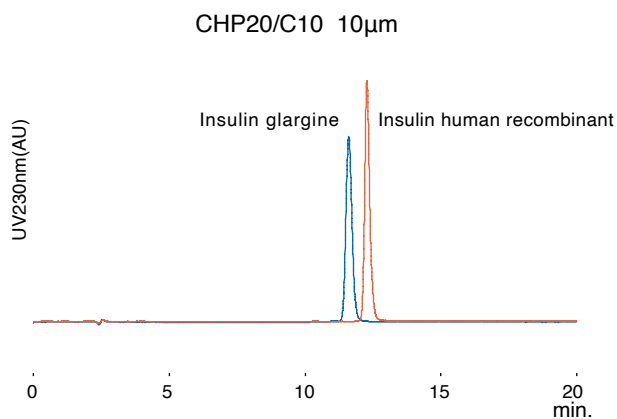


Conditions  
 Column : 150 ×4.6mm I.D.  
 Eluent : A :0.1% TFA;  
 B :0.1% TFA in AcCN  
 Flow rate : 1.00mL/min  
 Column temp. : 20%B-60%B over 20min;  
 Detection : UV280nm;  
 Sample : Ribonuclease A and Conalbumin 2mg/ml;  
 Injection : 10μL

# Application data of CHP series

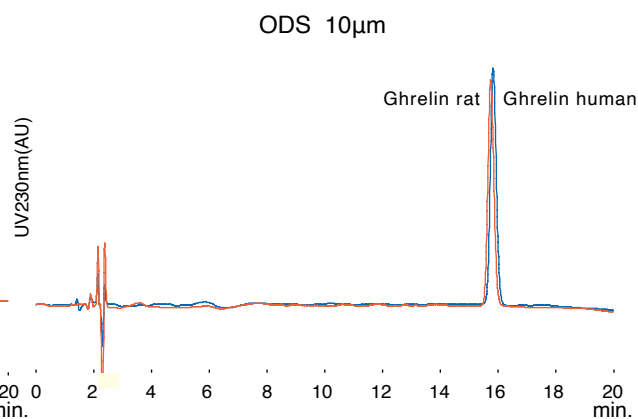
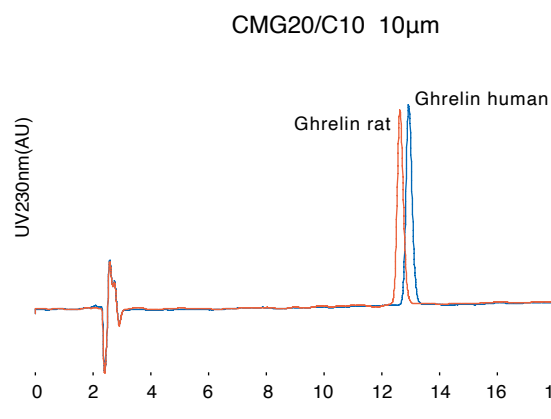
**Fig. 5-20 Insulin**

Conditions  
 Column : MCI GEL™ CHP20/C10  
 MCI GEL™ CMG20/C10  
 ODS 10μm  
 4.6mm I.D.×150mm  
 Eluent : A) 0.1%TFA, H<sub>2</sub>O  
 B) 0.1%TFA, CH<sub>3</sub>OH  
 Gradient : 20%B→60%B over 20min.  
 Flow rate : 1.0mL/min  
 Column temp. : 40°C  
 Detection : 280nm  
 Sample : Insulin Glargine and human recombinant , 1mg/mL each  
 Injection : 10μL



**Fig. 5-21 Ghrelin**

Conditions  
 Column : MCI GEL™ CMG20/C10  
 ODS 10μm  
 4.6mm I.D.×150mm  
 Eluent : A) 0.1%TFA, H<sub>2</sub>O  
 B) 0.1%TFA, AcCN  
 Gradient : 10%B→60%B over 25min.  
 Flow rate : 1.0mL/min  
 Column temp. : 40°C  
 Detection : 230nm  
 Sample : Ghrelin rat and Ghrelin human ,0.1mmol/l each  
 Injection : 10μL



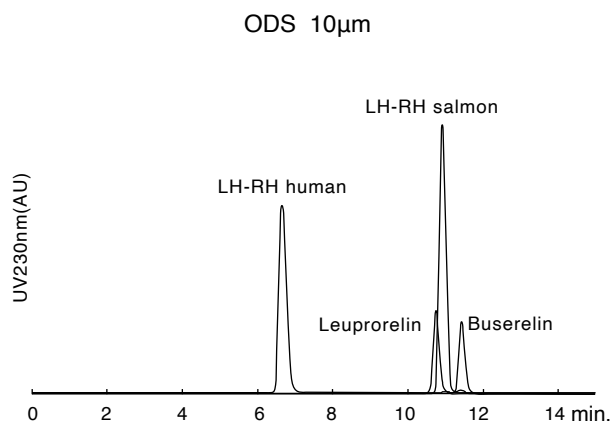
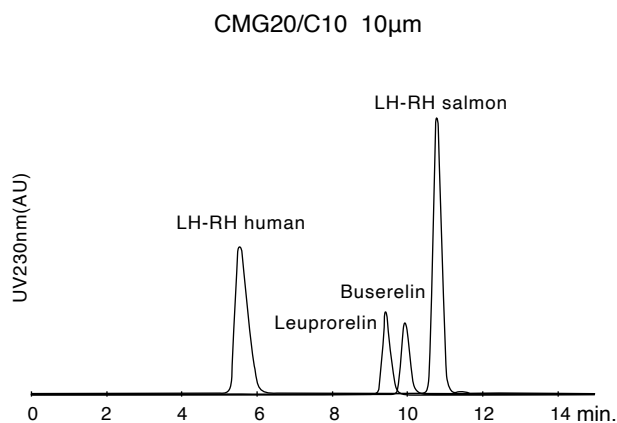
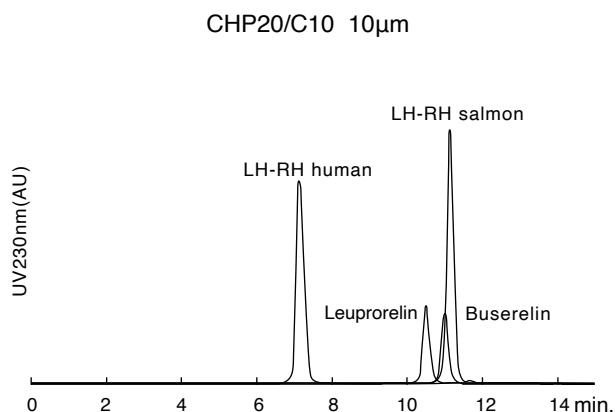
Column selection guide  
 1  
 Ion exchange columns and materials  
 2  
 Ion chromatography columns and materials  
 3  
 Bioseparation columns and materials  
 4  
 Analytical and preparative chromatography for pharmaceutical applications  
 5  
 Critical separation columns  
 6  
 SPE sorbent series  
 7  
 MCI GEL™ column list  
 8  
 MCI GEL™ material list  
 9  
 Compounds index  
 10

# Application data of CHP series

**Fig. 5-22 Leuprorelin**

**Conditions**

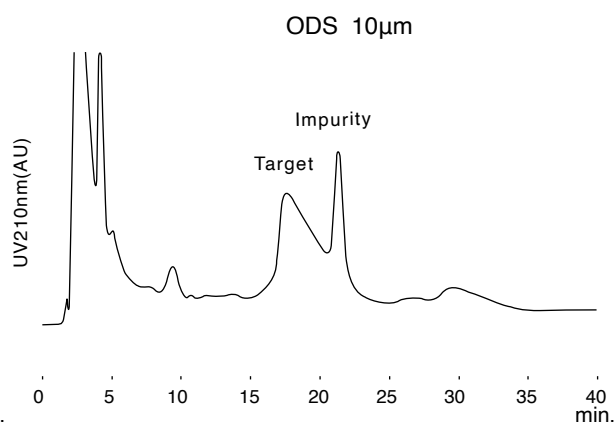
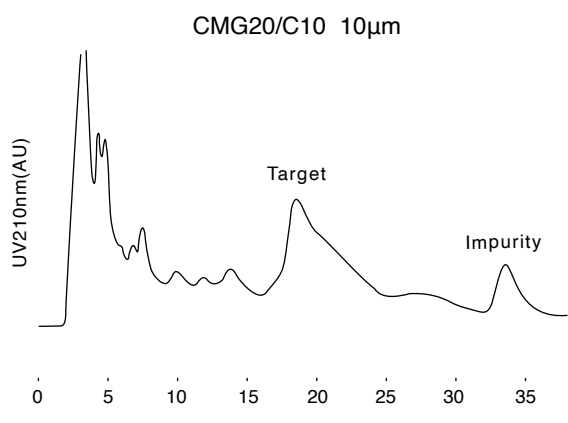
Column : MCI GEL™ CHP20/C10  
 MCI GEL™ CMG20/C10  
 ODS 10 $\mu$ m  
 4.6mm I.D.×150mm  
 Eluent : A) 0.1%TFA, H<sub>2</sub>O  
 B) 0.1%TFA, AcCN  
 Gradient : 20%B→60%B over 20min.  
 Flow rate : 1.0mL/min  
 Column temp.: 40°C  
 Detection : 280nm  
 Sample : Leuprorelin, LHRH human, LHRH salmon and Buserelin ,1mg/mL each  
 Injection : 10 $\mu$ L



**Fig. 5-23 Sifuvirtide**

**Conditions**

Column : MCI GEL™ CMG20/C10  
 ODS 10 $\mu$ m  
 4.6mm I.D.×150mm  
 Eluent : 0.1%TFA,/CH<sub>3</sub>CN=68/32  
 Flow rate : 1.0mL/min  
 Column temp.: 40°C  
 Detection : 210nm  
 Sample : Sifuvirtide crude(purity 35.5%) 2.1mg/mL  
 Injection : 0.4mL

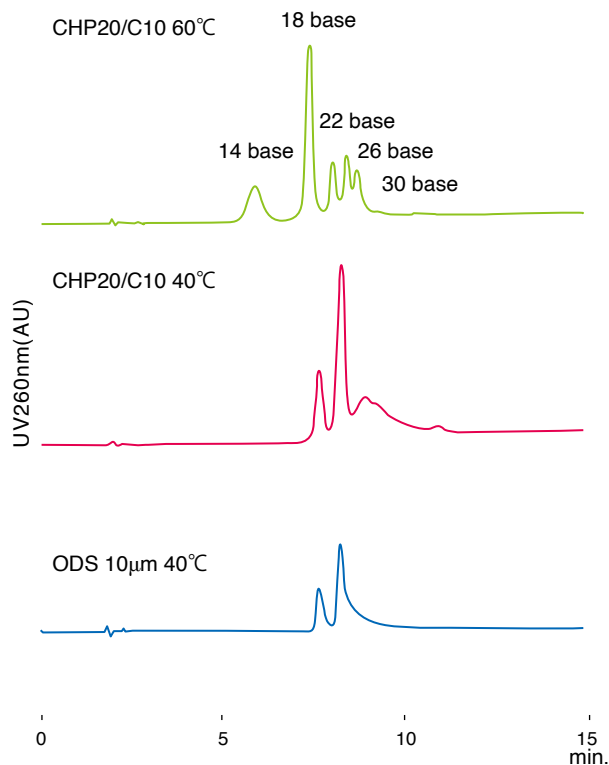


- 1 Column selection guide
- 2 Ion exchange columns and materials
- 3 Ion chromatography columns and materials
- 4 Bioprocession columns and materials
- 5 Analytical and preparative for pharmaceutical applications
- 6 Chiral separation columns
- 7 SPE sorbent series
- 8 MCI GEL™ column list
- 9 MCI GEL™ material list
- 10 Compounds index

# Application data of CHP series

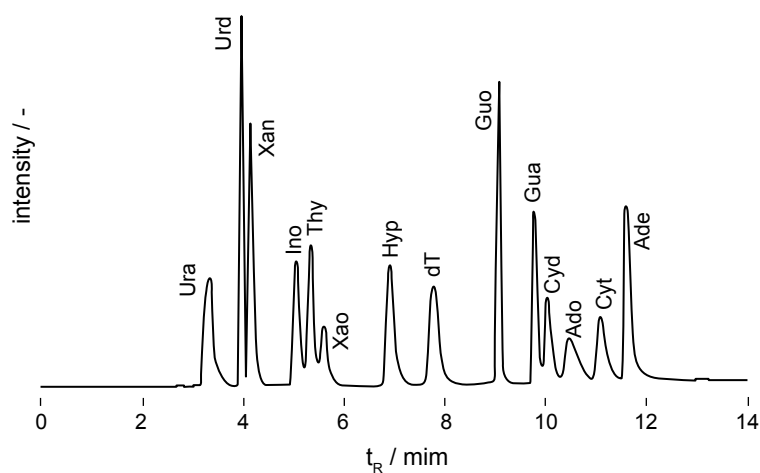
**Fig. 5-24 ssRNA Ladder Marker**

Conditions  
 Column : MCI GEL™ CMG20/C10  
 ODS 10 $\mu$ m  
 4.6mm I.D.×150mm  
 Eluent : A)100mM TEAA, H<sub>2</sub>O  
 B)100mM TEAA, CH<sub>3</sub>CN  
 Gradient : CHP10/C10 10%B→40%B over 30min  
 ODS 10 $\mu$ m 8%B→40%B over 30min  
 Flow rate : 1.0mL/min  
 Column temp.: 40°C  
 Detection : 260nm  
 Sample : 14-30 ssRNA Ladder Marker [max.0.04mg/mL]  
 Injection : 5 $\mu$ L



**Fig. 5-25 Nucleotide**

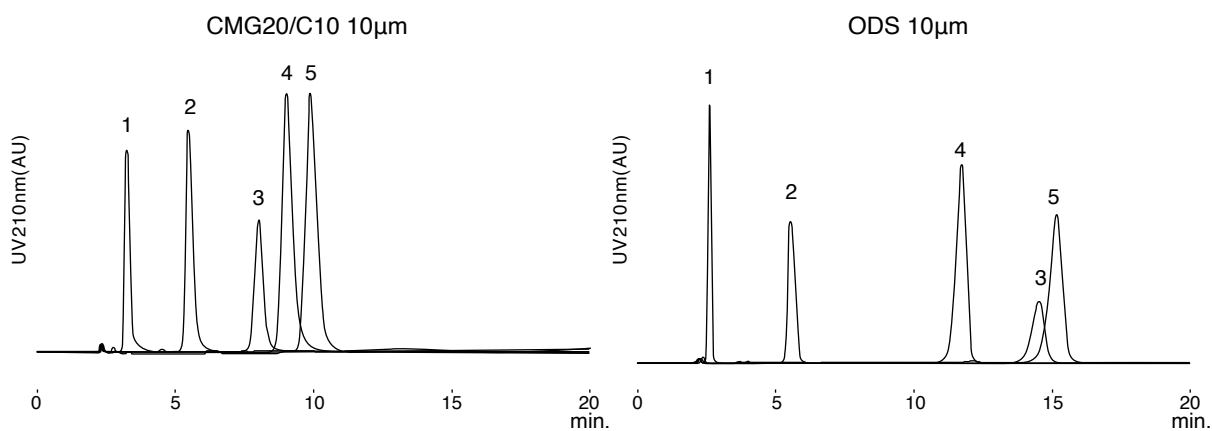
Conditions  
 Column : MCI GEL™ CHK40/C04  
 4.6mm I.D.×150mm  
 Eluent : A)19 mM H<sub>3</sub>PO<sub>4</sub> / 1 mM NaH<sub>2</sub>PO<sub>4</sub> / 5.0% ACN  
 B)20 mM Na<sub>2</sub>HPO<sub>4</sub> / 100 mM NaClO<sub>4</sub> / 30% ACN  
 Gradient : 0-4.0min 0%B 4.0-5.0min 0→30%B 5.0min-6.0min 30%B 6.0min-7.0min 30→50%B  
 7.0min-10.0min 50→65%B 10.0min-11.0min 65%B 11.0min- 0%B  
 Flow rate : 0.8mL/min  
 Column temp.: 50°C  
 Detection : UV260nm  
 Sample : 1.Ura, 2.Xan, 3.Thy, 4.Hyp, 5.Gua, 6.Cyt, 7.Ade, 8.Urd, 9.Xao, 10.dT, 11.Ino, 12.Guo, 13.Cyd, 14.Ado  
 Injection : 20 $\mu$ L



(Data provided by Professor Yokoyama of Yokohama National University)

# Application data of CHP series

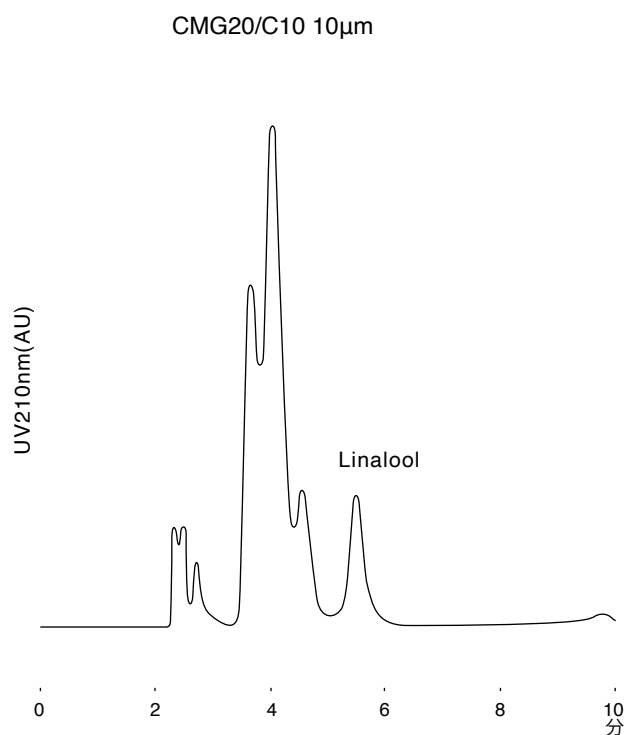
**Fig. 5-26 Linalool**



Conditions  
 Column : MCI GEL™ CMG20/C10  
           ODS 10µm  
           4.6mm I.D.×150mm  
 Eluent : Hexan/Ethanol=99.5/0.5  
 Flow rate : 1.0mL/min  
 Column temp.: 40°C  
 Detection : 210nm  
 Sample : 1:Linalyl Acrylate 1mg/mL  
           2:Linalool 1mg/mL  
           3:β-Citronellol 1mg/mL  
           4:Nerol 0.5mg/mL  
           5:Geraniol 0.5mg/mL  
 Injection : 10µL

**Fig. 5-27 Coriander**

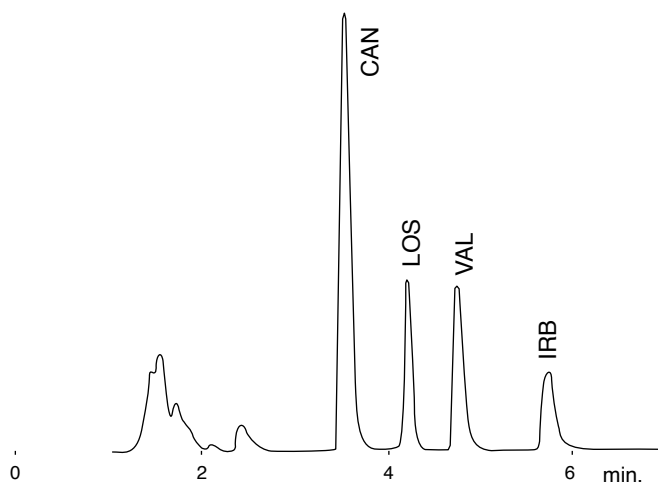
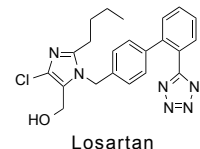
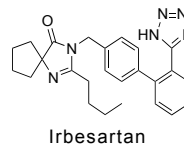
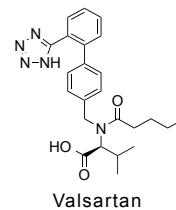
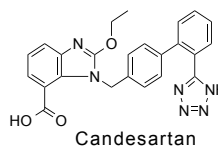
Conditions  
 Column : MCI GEL™ CMG20/C10  
           4.6mm I.D.×150mm  
 Eluent : Hexan/Ethanol=99.5/0.5  
 Flow rate : 1.0mL/min  
 Column temp.: 40°C  
 Detection : 210nm  
 Sample : Coriander  
 Injection : 10µL



# Application data of CHP series

**Fig. 5-28 Application data of CHK40/C04 : Sepatation of Sartans**

Conditions  
 Column : MCI GEL™ CHK40/C04  
 4.6mm I.D.×150mm  
 Eluent : A) 10 mM NaH<sub>2</sub>PO<sub>4</sub> +0.2 mM Na<sub>2</sub>HPO<sub>4</sub> (25%ACN)  
 B) 10 mM NaH<sub>2</sub>PO<sub>4</sub> +1.0 mM Na<sub>2</sub>HPO<sub>4</sub> (40%ACN)  
 Gradient : 0.5min 0%B 0.5-2.0min 50%B  
 2.0min-- 90%B  
 Flow rate : 1.0mL/min  
 Column temp. : 50°C  
 Detection : UV  
 Sample : Candesartan(CAN),Losartan(LOS),  
 Valsartan(VAL), Irbesartan(IRB)  
 Injection : 20μL

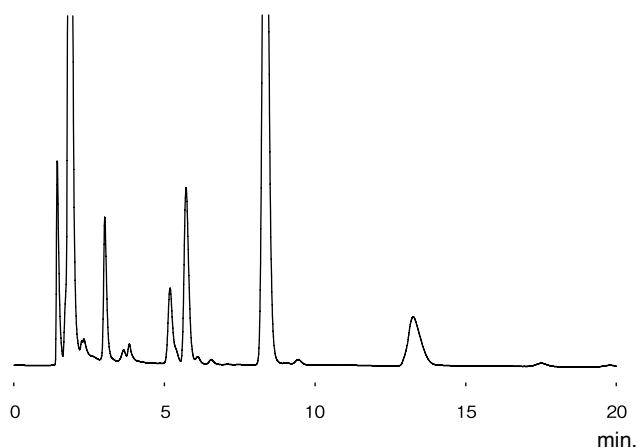


(Data provided by Professor Yokoyama of Yokohama National University)

## (Polyphenon 60)

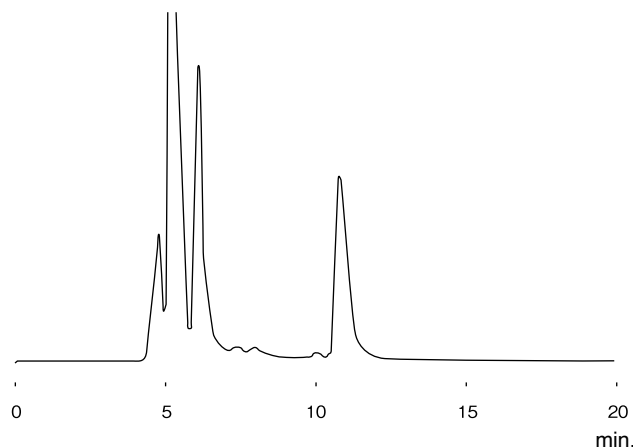
**Fig. 5-29 Modified Styrene Divinylbenzene  
CHP07/C04**

Conditions  
 Column : MCI GEL™ CHP07/C04  
 4.6mm I.D.×150mm  
 Eluent : CH<sub>3</sub>OH/10mM-Acetic acid=60/40  
 Flow rate : 0.46mL/min  
 Column temp. : 60°C  
 Detection : 280nm  
 Sample : Polyphenon 60(10mg/mL) each 10μL



**Fig. 5-30 Styrene Divinylbenzene CHP20/C04**

Conditions  
 Column : MCI GEL™ CHP20/C04  
 4.6mm I.D.×150mm  
 Eluent : CH<sub>3</sub>OH/10mM-Acetic acid=60/40  
 Flow rate : 0.46mL/min  
 Column temp. : 60°C  
 Detection : 280nm  
 Sample : Polyphenon 60(10mg/mL) each 10μL

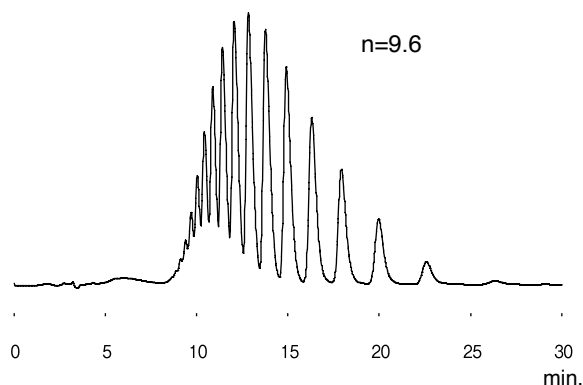
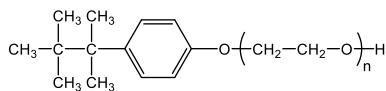


# Application data of CHP series

(TritonX-100)

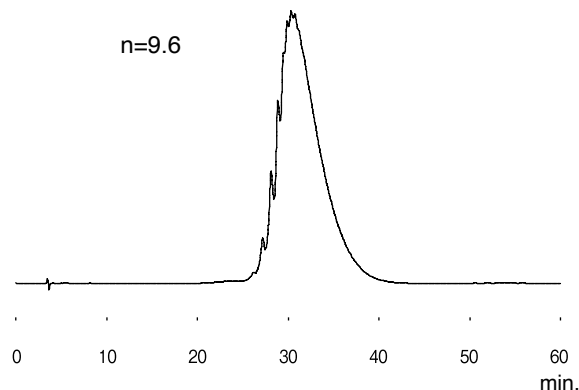
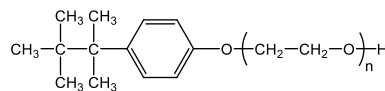
**Fig. 5-31 C18-alkylated aliphatics CHPOD/C04**

Conditions  
 Column : MCI GEL™ CHPOD/C04  
 4.6mm I.D.×150mm  
 Eluent : 50vol%CH<sub>3</sub>CN  
 Flow rate : 0.50mL/min  
 Column temp.: 40°C  
 Detection : 254nm  
 Sample : Triton X-100  
 (Polyoxyethylene octyl phenyl ether)  
 1% each 10μL



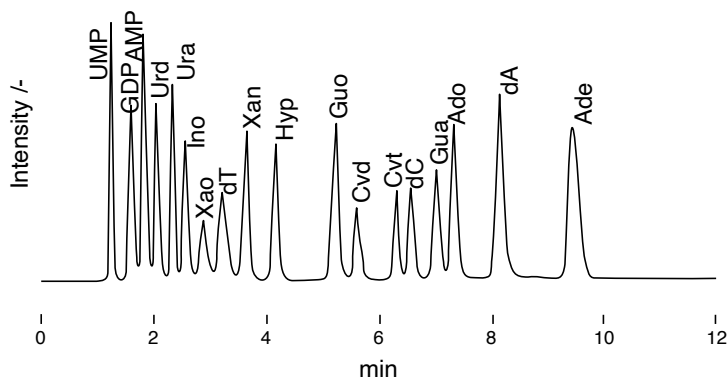
**Fig. 5-32 ODS (5μm)**

Conditions  
 Column : ODS (5μm)  
 4.6mm I.D.×250mm  
 Eluent : 50vol%CH<sub>3</sub>CN  
 Flow rate : 1.00mL/min  
 Column temp.: 40°C  
 Detection : 254nm  
 Sample : Triton X-100  
 (Polyoxyethylene octyl phenyl ether)  
 1% each 10μL

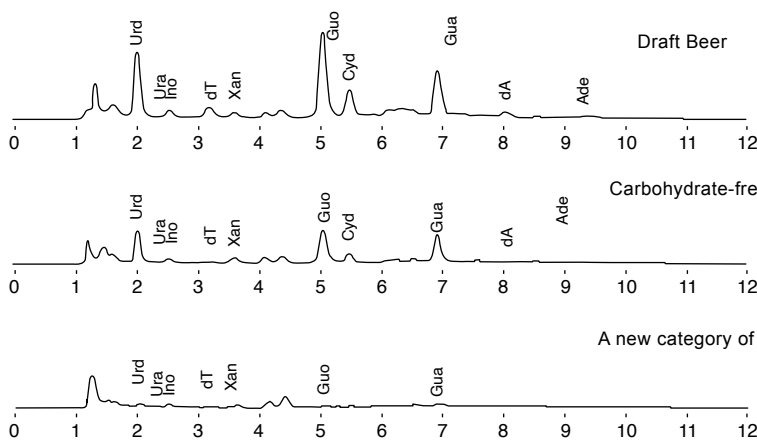


**Fig. 5-33 Application data of Nucleic base/Nucleoside and Beer**

Conditions  
 Column : MCI GEL™ CHK45/C05  
 4.6mm I.D.×150mm  
 Eluent : A) 8 mM H<sub>3</sub>PO<sub>4</sub>  
 B) 10 mM H<sub>3</sub>PO<sub>4</sub> /30% ACN  
 Gradient : 0-0.7min 0%B 0.7-3.0min 0→40%B 3.0-3.2min 40%B  
 3.2-3.5min 40→80%B 3.5-8.0min 80%B 8.0min-- 0%B  
 Flow rate : 1.3mL/min  
 Column temp.: 45°C  
 Detection : UV260nm  
 Injection : 20μL



Analysis of various category beer



(Data provided by Professor Yokoyama of Yokohama National University)